The over-expression of GH/GHR in tumour tissues with respect to healthy ones confirms its oncogenic role and the consequent oncosuppressor role of its physiological inhibitor, somatostatin: a review of the literature

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Abstract

The interaction between pituitary hormones, GH - PRL, and Growth Factors, GF, plays a fundamental role in the physiological and neoplastic mechanisms of growth, the latter using these factors to a much greater extent compared to the former, with a direct dose-dependent effect on the speed of local or metastatic expansion. In hormone-dependent tumours, the respective male and female sex hormones interact with GH - PRL - GF to sustain the expansion of the tumour. We carried out a review of the literature on the relationship between the expression of GH and GHR in tumour tissues compared to healthy tissues, and on the correlation between this expression and tumour aggressiveness. An over-expression of GH and GHR in tumours was a constant finding. In more than a thousand cases published in various clinical, observational, retrospective studies investigating cervico-facial tumours, lymphoproliferative diseases, breast cancer, prostate cancer, non-small-cell lung cancer, neuroblastomas, oesophageal cancer, glioblastomas, and sarcomas, we constantly found an improvement in objective response, quality of life and survival, compared to conventional oncological protocols, by inhibiting GH and correlated GF using somatostatin.

Abbreaviations:

GH	- Growth Hormone	Pit-1	- Pituitary-specific positive transcription factor 1
GHR	- Growth Hormone Receptor	hTERT	- human Telomerase Reverse Transcriptase
PRL	- Prolactin	EMT	- Epithelial Mesenchymal Transition
GF	- Growth Factor	MMP	- Metallo Proteases
IGF1	- Insulin Like Growth Factor	TSP1	- Thrombospondin 1
VEGF	- Vascular Endothelial Growth Factor	eNOS	- endothelial Nitric Oxide Synthesis
EGF	- Epidermal Growth Factor	bFGF	- basic Fibroblast Growth Factor
MAPK	- Mitogen-activated protein kinase	FSH	- Follicle-stimulating hormone
PI3K	- Phosphoinositide 3-kinase	LH	- Luteinizing hormone

INTRODUCTION

Endocrine, biological and biochemical data show the evident primary role of the GH-IGF1 axis in synergy with the other GH correlated growth factors (Fig.1), such as VEGF-A and EGF, together with prolactin (Fig.2). In hormone-dependent tumours, the respective sex hormones, oestrogen and testosterone, interact with GH-PRL-GF. GH is a peptide hormone consisting of 191 amino acids with a weight of 22.005 Da, synthesized, accumulated and secreted by the adenohypophysis. The numerous functions of GH include:

- Regulation of body growth
- Regulation of cell proliferation and differentiation

- Regulation of the metabolism of proteins, lipids and carbohydrates
- Increased protein synthesis in cells with multiple mechanisms including:
- Activation of some carriers of plasma membrane amino acids, causing increased entry into the cytoplasm.
- Transduction of cellular mRNA even without a greater than normal concentration of amino acids
- Increased protein synthesis by ribosomes

GH is the main mediator of the postnatal growth of somatic cells (Le Roith *et al.* 2001), and its effects on cell growth and differentiation are mediated through

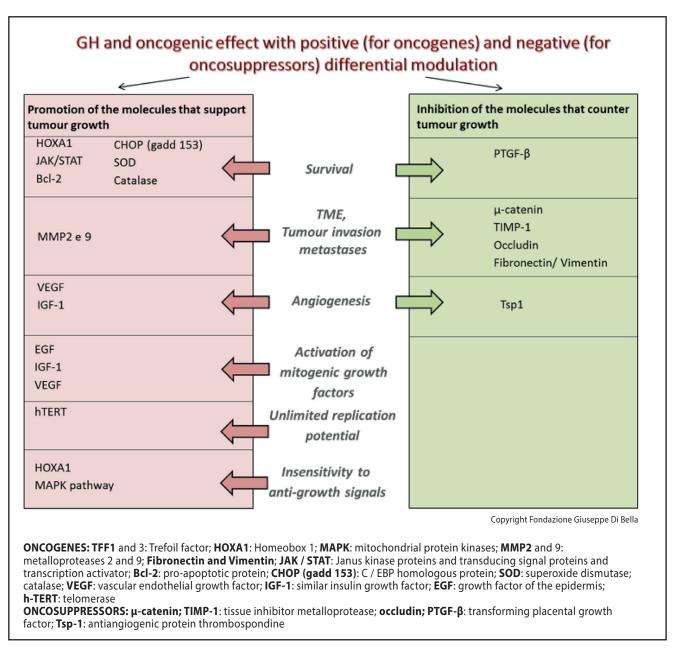


Fig. 1. Growth hormone interacts with many genes (oncogenes and oncosuppressors) that are part of growth different mechanism. At the middle of image (survival, TME, angiogenesis...) different mechanism of growth; in red factors and enzymes that are promoted by GH to induce tumour; in green factors that are inhibited by GH.

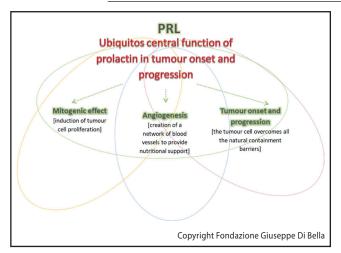


Fig. 2. Prolactin has a central role in tumour onset and progression through mitogenic effect and angiogenesis also.

interaction with its receptor (GH) (Le Roith *et al.* 2001; Zhu *et al.* 2001) which activates signal transduction pathways critical for cell growth and survival including signal transductors (JAK-2/STAT), the cascade of the mitogen-activated protein kinase (P44/42 (MAPK), and of the phosphoinositide 3-kinase (PI3K) (Le Roith *et al.* 2001; Zhu *et al.* 2001). The receptor of GH belongs to the large family of cytokine or hematopoietic receptors, which do not contain the tyrosine-kinase domain in their cytoplasm region.

The family of the GH/cytokine receptors (Class I) and the family of the interferon receptors (Class II) share structural characteristics, recently identified, and common signal transduction pathways. In addition, both classes of receptors are associated with various members of the family of Janus tyrosine kinases and activate a new family of transcription factors, known as transcription transducers and activators, which relate ligands to activation of gene expression (Goffin & Kelly, 1996).

To understand the actions of GH it is necessary to know the structure and expression of its receptor. The cloning and sequencing of the GH receptor showed that this receptor is not homologous to other receptors with a known function, but the GH – Prolactin's receptor homology is a scientifically and clinically important factor due to its therapeutic repercussions. A widespread distribution with variable concentrations of GHR was observed in many types of normal and tumour cells, with a marked and significant prevalence in tumour cells in proportion to the proliferative index and to the invasive and metastasizing capacity (Lincoln et al. 1998).

The expression of the IGF-1 gene is regulated differently by the GH in different tissues (Lowe *et al.* 1987). The dependence on GH of IGF-1 has been extensively confirmed in humans. In patients with complete GH deficiency, the levels of IGF-1 are always decreased, with the lowest levels found in patients with Laron

dwarfism in whom the GHR is missing (Daughaday & Trivedi 1987) The administration of GH causes an increase of IGF-1 in GH-responsive patients. However, the co-localization of IGF-1 and GHR is not necessary since the role of GH in the tumour cells may be to regulate the function of the mature cells rather than to promote cell proliferation by local synthesis of IGF-1. These two hormones do not always act in harmony; in some tissues the synthesis of IGF-1 is independent of GH (Hynes et al. 1987), despite the fact that these tissues possess GHR, as shown by a powerful mitogenesis independent of IGF-1 in response to GH (Rabinovitch et al. 1983). More in-depth studies are required to establish whether, and in what conditions and in which tissues GH can act independently of the synthesis of IGF-1 in human tumour cells.

A high concentration of the receptor was also constantly observed in cell lines from tissue in the exponential growth stage. The protein that binds the GH in the nucleus has the same binding sites as cytoplasmic and membrane GHR. The GH / IGF-1 axis is the main mediator of somatic growth and during infancy plays an essential role in the development of the mammary gland, regulating cell proliferations, differentiation and apoptosis (Kleinberg 1997). The determining role of the GH / IGF-1 axis (Laban *et al.* 2003), and its close receptor, functional, and proliferative interaction with the other pituitary secretion, prolactin, with other GH-correlated growth factors, such as VEGF and EGF, is therefore scientific evidence in tumoral genesis.

Review of the clinical and experimental studies confirming the mitogenic properties of GH and the relative biochemical and molecular mechanisms of action

IN BREAST CANCER

- The expression of GHR is associated with the malignant transformation of the mammary gland in a dose-dependent manner (Lincoln et al. 1998);
- Human recombinant GH increases proliferation in breast cancer (Conte *et al.* 1990);
- There is an increase in incidence and aggressiveness of secondary malignant tumours, including breast cancer, in patients on the Childhood Cancer Survivor Study treated with rhGH to maximize growth (Rutter & Rose 2007; Neglia et al. 2001; Ergun-Longmire et al. 2006; Sklar et al. 2002);
- IGF-1, the mitogen that acts as an intermediary in the action of GH, is over-expressed in breast cancer and the bond with IGF-1 is higher in breast cancer tissue than in normal adjacent tissue; (Arteaga & Osborne 1989; Yee *et al.* 1989)
- The expression of the protein and of mRNA of GHR has been identified in human breast cancer (Mertani *et al.* 1998; Decouvelaere *et al.* 1995);
- Cell lines of breast cancer produce and secrete IGF-1 (Huff *et al.* 1986);

- Human recombinant GH increases proliferation in breast cancer (Conte et al. 1990);
- An increase in levels of HGHR mRNA in samples of human breast cancer with respect to adjacent normal tissue (Gebre-Medhin et al. 2001);
- RT-PCR and Western blot analysis showed the expression of GHRH and its receptor in breast cancer, and antagonists of GHRH inhibit the growth of tumors (Schally &Varga 2006);
- The proteins that regulate the secretion of hGH by the hypophysis are implicated in breast cancer (Chatzistamou et al. 2004; Kahán et al. 1999);
- The induction at the forced expression of Pit-1 increases the expression of GH mRNA and proliferation in human breast cancer cells (Gil-Puig et al. 2005);
- GH favours the immortalization of mammary epithelial cell lines through the increase of the levels of mRNA and of proteins of catalytic subunit of telomerase, hTERT (Emerald *et al.* 2007; Dimri *et al.* 2005; Stewart &Weinberg 2006);
- autocrine GH inhibits the mechanisms of anchorage and adhesion in breast cancer cells and of tumour growth in vitro (Kaulsay et al. 1999; Mukhina et al. 2004);
- the expression of autocrine hGH therefore satisfies the criteria of being considered as an oncogene for human breast cancer (Lincoln *et al.* 1998);
- the expression of GH in MCF-10A cells leads to filling of the lumen due to upset of the normal mammary gland architecture and of the proliferative order (Zhu et al. 2001);
- GH increases the metastasizing of breast cancer by interruption of the cell-cell contact and increase in cell migration and invasion (Mukhina et al. 2004);
- IGF-1 (GH-correlated) is over-expressed in breast cancer (Yee *et al.* 1989);
- GH is higher in breast cancer tissue with respect to adjacent normal tissue (Arteaga &Osborne 1989);
- Recent studies showed that the expression of GH can increase telomerase activity and extend the replicative ability of a primary mammary epithelial cell line (Emerald *et al.* 2007);

IN GENERAL IN TUMORS

- GH significantly increases the expression of the proto-oncogene c-myc (Murphy *et al.* 1987);
- At high concentrations, GH directly accelerates the growth of osteogenic sarcoma, at the same time inducing high levels of somatomedin (Ward et al. 1987; Ratner & Hare 1983);
- GH accelerates the growth of multiple myelomas (Hägg et al. 1988);
- the GHI/GF1 axis is highly represented in human lung cancer tissue obtained immediately after surgery, compared to healthy surrounding lung tissue (Minuto et al. 1986);

- high plasma levels of GH are documented in numerous human tumours (Adamson *et al.* 1980; Andrews 1983);
- an increased concentration of GH was demonstrated in bone tumours (Ratner & Hare 1983);
- a high and significant concentration of GH was found in multiple myelomas (Hägg *et al.* 1988);
- in human lung cancer tissue obtained immediately after surgery, GH-dependent concentrations of IGF-1 GH were found to be decidedly higher than in the normal surrounding tissue (Minuto *et al.* 1986);
- an increase in incidence and aggressiveness of Hodgkin's lymphoma and colon-rectal cancer was seen in patients treated with hGH during infancy or early adulthood (Swerdlow et al. 2002);
- IGF-1 is over-expressed in tumours of the colon (Sklar *et al.* 2002);
- Various clinical studies and case reports have shown an increased incidence of polyps, adenomatosis of the colon and cancer of the colon in patients with acromegaly (Ron et al. 1991; Ziel & Peters 1988; Pines et al. 1985; Brunner et al. 1990);
- The expression of GHR was shown in cell membrane, cytoplasm, the nucleus of normal tissues and in greater concentrations in tumour cells (Lincoln *et al.* 1998);
- The expression of autocrine GH promotes cell proliferation (Kaulsay *et al.* 1999);
- Autocrine GH is the first example of a human gene that can both potentially immortalize and oncogenically transform the human epithelial cell (Lincoln *et al.* 1998);
- GH promotes the phenotypic conversion of cells from epithelial to mesenchymal morphology (EMT) with the acquisition of a migratory and invasive epithelial-mesenchymal phenotype, through the down regulation of plakoglobin, re-localization of E-cadherin to the cytoplasm, and a greater activity of matrix metalloproteases 2 and 9 (MMP) (Mukhina et al. 2004; Sommers et al. 1994; Thiery 2002);
- GH promotes the migration of endothelial cells and angiogenesis, and increases the levels of VEGF-A mRNA and (Brunet-Dunand *et al.* 2009) (Fig. 3) by down-regulation of thrombospondin 1 (TSP1), inhibitor of the angiogenic phenotype (Lawler & Detmar 2004);
- The transduction of GH leads to a significant induction of numerous genes of angiogenic inductors such as endothelial nitric oxide synthesis (eNOS), and of angiogenic growth factors such as VEGF basic fibroblast growth factor (bFGF), while immunohistochemical analysis revealed an increase in capillary density and cell proliferation (Kusano et al. 2007);
- the GH/IGF-1 axis has a protective effect against radiation-induced programmed cell death (Jameel et al. 2004; Perry et al. 2006);
- GH inhibits apoptosis pathways, negatively regulating cell growth stop genes such as gadd153 / CHOP

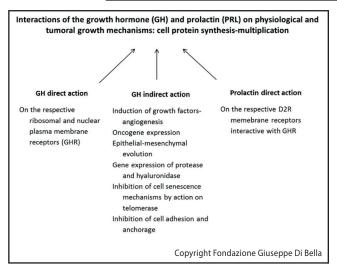


Fig. 3. Cooperation between direct and indirect actions of GH and prolactin on tumour induction.

(C / EBP), thus protecting tumour cells also from oxidative stress (Zhu *et al.* 2005)

- GH is radioprotective in various breast cancer cell lines (Brunet-Dunand et al. 2009);
- In addition, the over-expression of HGHR mRNA and of proteins predicts the response to radiotherapy in rectal cancer (Wu et al. 2006);

The concept of the binary proliferative GH/IGF1 axis can be extended to the quaternary GH/IGF1/PRL/ ER axis in breast cancer (Fig.3). The mitogenic synergism of the growth hormone with the Insulin-like growth factor, with Prolactin and testosterone in male prostate cancer and with oestrogens in female reproduction system tumours and breast cancer has now in fact been confirmed (Gallego et al. 2001). We believe it is useful to highlight the fact that most of the mitogenic effects of GH in somatic cells are mediated not only by hepatic IGF-1 but also by induction of the expression of other growth factors such as EGF (Vacas et al. 2016) and VEGF-A (Brunet-Dunand et al. 2009). The bond between GH and GHR, as well as the documented positive regulation of IGF1, EGF and VEGF, activates the signal transduction of various pathways including JAK-2/STAT, MAPK, and PIK3.

It is a significant documented fact that subjects with acromegaly have an increased risk of colon-rectal cancer (Jenkins *et al.* 2006). High serum levels of GH (Emerman *et al.* 1985) and consequently of IGF-1 (Laban *et al.* 2003), (Yakar *et al.* 2005; Khandwala *et al.* 2000) have been observed in 40% of patients with breast cancer. In other tumours too, such as lymphoproliferative diseases, the expression of GHR has been observed, consistently with its presence in cultivated human lymphocytes (IM 9 cell line) shown with radio-marked ligands (Hughes & Friesen 1985). The GH/IGF-1 axis also modulates the immune system, although the way in which this interaction takes place

and the proportional relationships have not been clarified. GH directly regulates the function of the lymphocytes through its receptor (Lesniak *et al.* 1987), or with an action mediated by IGF-1 (Kozak *et al.* 1987). The increase of GHR in melanocytes, nevi, primary melanomas and metastatic melanomas is evidence of its activation of the tumour's progression in these diseases. The expression of GHR in benign prostate hyperplasia and in carcinoma of the prostate is proportional, with a dose-dependent relationship with the aggressiveness and proliferative index of these cell clones (El Etreby & Mahrous 1979; Sinowatz *et al.* 1991; Bengtsson *et al.* 1988).

Clinical, retrospective observational studies confirmed the evident anticancer efficacy of the inhibition of GH in breast cancer (Di Bella *et al.* 2013a), as in many other non-neuroendocrine tumours, such as sarcomas (Di Bella *et al.* 2015b), glioblastomas (Di Bella *et al.* 2015a), neuroblastomas (Di Bella & Colori 2009a), cervico-facial tumours (Di Bella *et al.* 2012a), oesophageal tumours (Di Bella &Madarena 2009), non-small cell lung cancer (Norsa & Martino 2006), chronic lymphatic leukaemia (Todisco . 2009; Di Bella *et al.* 2012), and Hodgkin and Non Hodgkin lymphoma (Todisco *et al.* 2001), by means of its physiological antidote, somatostatin.

CONCLUSIONS

These experimental and clinical data, consistent with the biological function of GH, provide further confirmation of the oncogenic induction of its over-expression and the dose-dependent relationship between the extent of the GH/IGF1/GHR expression and the proliferative and aggressive characteristics of the tumour clones (Wu et al. 2011). Through the differential regulation of the gene expression, autocrine GH also regulates vital molecular and biochemical mechanisms such as cell growth and survival, migration and invasion, epithelial-mesenchymal transition (EMT), replication potential and oncogenic transformation. The genes that autocrine GH regulates positively or negatively to induce oncogenesis are known. (Perry et al. 2008).

This scientific evidence therefore fully validates the rationale for the generalised use in oncotherapy of somatostatin which acts indifferently and equally on pineal and autocrine GH, regardless of the presence of SSTR in the tumour cells. Since GH and correlated growth factors are over-expressed in all tumours, albeit to different extents, with activation of numerous proliferative and angiogenic signalling pathways, the negative regulation of GH by means of somatostatin is logical, and is extended to the correlated growth factors, as widely documented in the literature (Fig.4).

The generalised anticancer use of somatostatin is therefore justified, since it antagonizes the common denominators, and causal factors of all tumours, and the over-expression of the Growth hormone – corre-

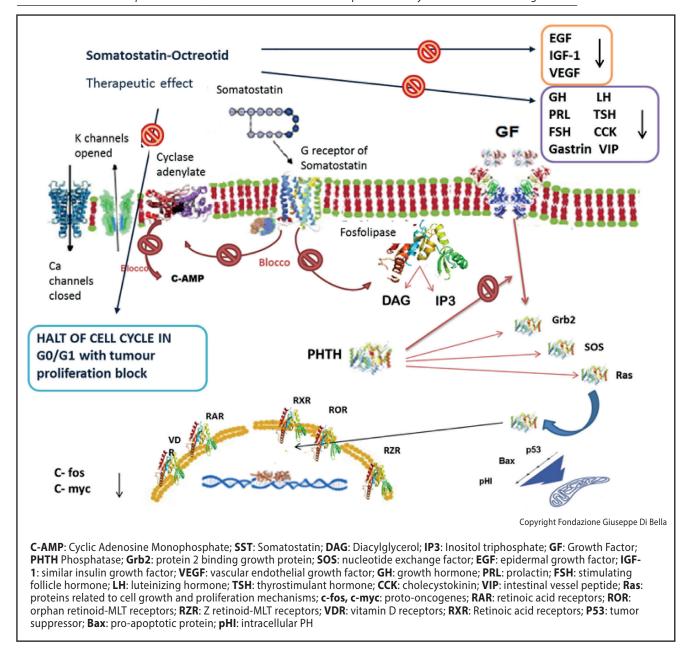
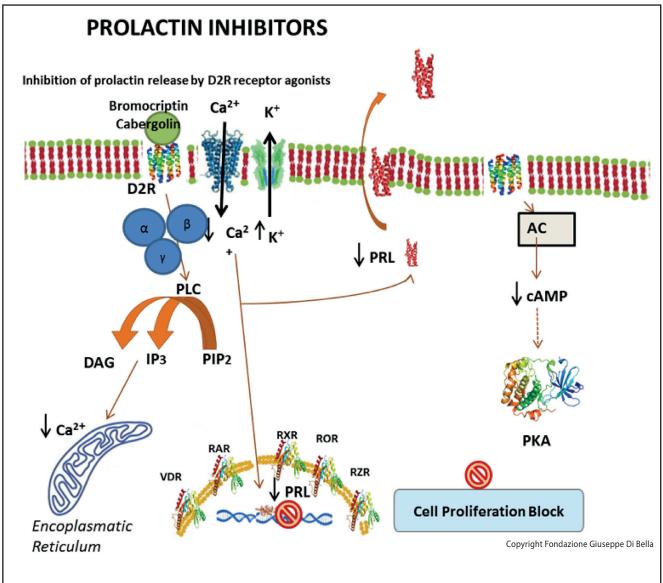


Fig. 4. Somatostatin's pathway (and its analogous, octreotid). These molecules block survival mechanisms (c-AMP, Fosfolipase, Growth Factors...) arresting cell cycle in G0/G1.

lated Growth Factors axis. In breast cancer, the close receptor interaction of GH with prolactin and the functional interaction with oestrogen lead, in a multitherapy setting, to synergic inhibition of GH-GF with somatostatin/octreotide (Fig.4), and of prolactin with D2R agonists (Fig.5), and estrogenic interaction by means of FSH-LH analogues and aromatase inhibitors, with decided progress in the treatment of these tumours (Di Bella *et al.* 2013b; Di Bella 2011). In cases of breast cancer monitored at 5 years, this biological multitherapy synergistically reinforced by the cytostatic, differentiating, immunomodulating and trophic functions of Melatonin, Solution of retinoids in vitamin E, vitamin D3 and vitamin C, (Di Bella Method), without toxic

effects, significantly improved quality of life, objective response and survival compared to the same tumour stages of breast cancer treated with conventional oncological protocols (Di Bella *et al.* 2013b; Di Bella *et al.* 2018; Di Bella 2011; Di Bella & Colori 2009b) . In the same way, in prostate cancer the synergic inhibition of GH-GF with somatostatin/octreotide, of prolactin with D2R agonists, and the androgenic block by means of bicalutamide and FSH-LH analogues allowed progress in the treatment of these tumours (Di Bella *et al.* 2013b). We draw attention to these concepts and these data with the intention of improving the prognosis of tumours by making use of this currently undervalued scientific evidence.



AC: Adenylate cyclase; C-AMP: Cyclic Adenosine Monophosphate; PKA: protein kinase A; PRL: prolactin; D2R: dopamine receptor 2; PLC: phospholipase; PIP2: phosphoinositol biphosphate; DAG: Diacylglycerol; IP3: Inositol triphosphate; RAR: retinoic acid receptors; ROR: orphan retinoid-MLT receptors; RZR: Z retinoid-MLT receptors; VDR: vitamin D receptors; RXR: Retinoic acid receptors

Fig. 5. Bromocriptin and cabergolin are inhibitors of prolactin' release, binding D2R receptor. The activation of D2R receptor induces different mechanism in cell until proliferation block.

REFERENCES

- 1 Adamson U, Broström LA, Efendić S, Hall K (1980). Glucose Tolerance, Growth Hormone and Somatomedin Levels in Osteosarcoma Patients. Acta Endocrinologica 94(4): 517–522. http://www.ncbi.nlm.nih.gov/pubmed/7001830.
- 2 Andrews GS (1983). Growth Hormone and Malignancy. Journal of Clinical Pathology 36 (8): 935–937. http://www.ncbi.nlm.nih.gov/ pubmed/6875020.
- 3 Arteaga CL, Osborne CK (1989). Growth inhibition of human breast cancer cells in vitro with an antibody against the type I somatomedin receptor. Cancer Res. 49(22): 6237-41. PubMed PMID: 2553250. http://cancerres.aacrjournals.org/content/ canres/49/22/6237.full.pdf.
- 4 Bengtsson BA, Edén S, Ernest I, Odén A, Sjögren B (1988). Epidemiology and Long-Term Survival in Acromegaly. A Study of 166 Cases Diagnosed between 1955 and 1984. Acta Med Scand 223 (4): 327–35. http://www.ncbi.nlm.nih.gov/pubmed/3369313.

- 5 Brunet-Dunand SE, Vouyovitch C, Araneda S, Pandey V, Vidal LJ, Print C et al. (2009). Autocrine human growth hormone promotes tumor angiogenesis in mammary carcinoma. Endocrinology. 150(3): 1341-1352. doi: 10.1210/en.2008-0608. Epub 2008 Oct 30. PubMed PMID: 18974274.
- 6 Brunner JE, Johnson CC, Zafar S, Peterson EL, Brunner JF, Mellinger RC (1990). Colon cancer and polyps in acromegaly: increased risk associated with family history of colon cancer. Clin Endocrinol (Oxf). 32(1): 65-71. PubMed PMID: 2331812.
- 7 Chatzistamou I, Schally AV, Kiaris H, Politi E, Varga J, Kanellis G, Kalofoutis A, Pafiti A, Koutselini H. I (2004) mmunohistochemical detection of GHRH and its receptor splice variant 1 in primary human breast cancers. Eur J Endocrinol. 151(3): 391-396. PubMed PMID: 15362970. http://www.eje-online.org/content/151/3/391. full.pdf.
- 8 Conte PF, Gardin G, Pronzato P, Miglietta L, Rosso R, Amadori D et al. (1990). In vivo manipulation of human breast cancer growth by estrogens and growth hormone: kinetic and clinical results. J Steroid Biochem Mol Biol. 37(6): 1103-1108. PubMed PMID: 1981014.

- 9 Daughaday WH, Trivedi B (1987). Absence of Serum Growth Hormone Binding Protein in Patients with Growth Hormone Receptor Deficiency (Laron Dwarfism). Proceedings of the National Academy of Sciences of the United States of America 84 (13). National Academy of Sciences: 4636–4640. http://www.ncbi.nlm.nih.gov/pubmed/3474620.
- 10 Decouvelaere C, Peyrat JP, Bonneterre J, Djiane J, Jammes H (1995). Presence of the two growth hormone receptor messenger RNA isoforms in human breast cancer. Cell Growth Differ. 6(4): 477-483. PubMed PMID: 7794815.
- 11 Di Bella G (2011). The Di Bella Method (DBM) Improved Survival, Objective Response and Performance Status in a Retrospective Observational Clinical Study on 122 Cases of Breast Cancer. Neuro Endocrinol Lett 32(6): 751–762. http://www.ncbi.nlm.nih. gov/pubmed/22167148.
- 12 Di Bella G, Mascia F (2013a). The Di Bella Method (DBM) in the Treatment of Prostate Cancer: A Preliminary Retrospective Study of 16 Patients and a Review of the Literature. Neuro Endocrinol Lett. **34**(6): 523-528.
- 13 Di Bella G, Colori B, Scanferlato R (2018). The Synergism of Somatostatin, Melatonin, Vitamins Prolactin and Estrogen Inhibitors Increased Survival, Objective Response and Performance Status In 297 Cases of Breast Cancer. Transl Biomed. 9(1): 144. DOI: 10.21767/2172-0479.100144
- 14 Di Bella G, Colori B (2009). Complete Objective Response of Neuroblastoma to Biological Treatment. Neuro Endocrinol Lett 30(4): 437–449. http://www.ncbi.nlm.nih.gov/pubmed/20010503.
- 15 Di Bella G, Colori B, Mascia F (2012). The Di Bella Method (DBM) Improved Survival, Objective Response and Performance Status in a Retrospective Observational Clinical Study on 55 Cases of Lymphomas. Neuro Endocrinol Lett **33**(8): 773–781. http://www.ncbi.nlm.nih.gov/pubmed/23391973.
- 16 Di Bella G, Leci J, Ricchi A, and Toscano R (2015a). Recurrent Glio-blastoma Multiforme (Grade IV WHO 2007): A Case of Complete Objective Response Concomitant Administration of Somatostatin / Octreotide, Retinoids, Vit E, Vit D3, Vit C, Melatonin, D2 R Agonists (Di Bella Method. Neuro Endocrinol Lett 36(2): 127–32. http://www.ncbi.nlm.nih.gov/pubmed/26071580.
- 17 Di Bella G, Madarena M (2009). Complete Objective Response of Oesophageal Squamocellular Carcinoma to Biological Treatment. Neuro Endocrinol Lett **30**(3): 312–321. http://www.ncbi.nlm.nih.gov/pubmed/19855352.
- 18 Di Bella G, Mascia F, Ricchi A, Colori B (2013b). Evaluation of the Safety and Efficacy of the First-Line Treatment with Somatostatin Combined with Melatonin, Retinoids, Vitamin D3, and Low Doses of Cyclophosphamide in 20 Cases of Breast Cancer: A Preliminary Report. Neuro Endocrinol Lett 34 (7): 660–668. http://www.ncbi. nlm.nih.gov/pubmed/24464005.
- 19 Di Bella G, Toscano R, Ricchi A, Colori B (2015b). Congenital Fibrosarcoma in Complete Remission with Somatostatin, Bromocriptine, Retinoids, Vitamin D3, Vitamin E, Vitamin C, Melatonin, Calcium, Chondroitin Sulfate Associated with Low Doses of Cyclophosphamide in a 14-Year Follow Up. Neuro Endocrinol Lett 36(8): 725–733. http://www.ncbi.nlm.nih.gov/pubmed/26921571.
- 20 Dimri G, Band H, Band V (2005). Mammary epithelial cell transformation: insights from cell culture and mouse models. Breast Cancer Res. 7(4): 171-179. Epub 2005 Jun 3. Review. PubMed PMID: 15987472; PubMed Central PMCID: PMC1175079.
- 21 Emerald BS, Chen Y, Zhu T, Zhu Z, Lee KO, Gluckman PD et al. (2007). AlphaCP1 mediates stabilization of hTERT mRNA by autocrine human growth hormone. J Biol Chem. **282**(1): 680-690. Epub 2006 Nov 3. PubMed PMID: 17085453.
- 22 Emerman JT, Leahy M, Gout PW, Bruchovsky N (1985). Elevated growth hormone levels in sera from breast cancer patients. Horm Metab Res. 17(8): 421-424. PubMed PMID: 4054832.
- 23 Ergun-Longmire B, Mertens AC, Mitby P, Qin J, Heller G, Shi W, Yasui Y, Robison LL, Sklar CA (2006). Growth hormone treatment and risk of second neoplasms in the childhood cancer survivor. J Clin Endocrinol Metab. 91(9): 3494-3498. Epub 2006 Jul 5. PubMed PMID: 16822820.

- 24 El Etreby MF, Mahrous AT (1979). Immunocytochemical Technique for Detection of Prolactin (PRL) and Growth Hormone (GH) in Hyperplastic and Neoplastic Lesions of Dog Prostate and Mammary Gland. Histochemistry **64** (3): 279–286. http://www.ncbi.nlm.nih.gov/pubmed/93102.
- 25 Gallego MI, Binart N, Robinson GW, Okagaki R, Coschigano KT, Perry J et al. (2001). Prolactin, growth hormone, and epidermal growth factor activate Stat5 in different compartments of mammary tissue and exert different and overlapping developmental effects. Dev Biol. 229(1): 163-175. PubMed PMID: 11133161.
- 26 Gebre-Medhin M, Kindblom LG, Wennbo H, Törnell J, Meis-Kindblom JM (2001). Growth hormone receptor is expressed in human breast cancer. Am J Pathol. 158(4): 1217-1222. PubMed PMID: 11290538; PubMed Central PMCID: PMC1891910.
- 27 Gil-Puig C, Seoane S, Blanco M, Macia M, Garcia-Caballero T, Segura C et al. (2005). Pit-1 is expressed in normal and tumorous human breast and regulates GH secretion and cell proliferation. Eur J Endocrinol. 153(2): 335-344. PubMed PMID: 16061841.
- 28 Goffin V, Kelly PA (1996). Prolactin and Growth Hormone Receptors. Clin Endocrinol 45(3): 247–255. http://www.ncbi.nlm.nih.gov/pubmed/8949560.
- 29 Hägg E, Asplund K, Holm J (1988). Acromegaly and Multiple Myeloma. Ann Intern Med **109**(5): 437–438.
- 30 Huff KK, Kaufman D, Gabbay KH, Spencer EM, Lippman ME, Dickson RB (1986). Secretion of an insulin-like growth factor-I-related protein by human breast cancer cells. Cancer Res. 46(9): 4613-4619. PubMed PMID: 3731113.
- 31 Hughes JP, Friesen HG (1985). The Nature and Regulation of the Receptors for Pituitary Growth Hormone. Annual Rev Physiol 47(1): 469–482. doi:10.1146/annurev.ph.47.030185.002345.
- 32 Hynes MA, Van Wyk JJ, Brooks PJ, D'Ercole AJ, Jansen M, Lund PK (1987). Growth hormone dependence of somatomedin-C/insulin-like growth factor-I and insulin-like growth factor-II messenger ribonucleic acids. Mol Endocrinol. (3): 233-242. PubMed PMID: 3453890.
- 33 Jameel JK, Rao VS, Cawkwell L, Drew PJ (2004). Radioresistance in carcinoma of the breast. Breast. 13(6): 452-460. Review. PubMed PMID: 15563851.
- 34 Jenkins PJ, Mukherjee A, Shalet SM (2006). Does Growth Hormone Cause Cancer? Clin Endocrinol 64(2): 115–21. doi:10.1111/j.1365-2265.2005.02404.x.
- 35 Kahán Z, Arencibia JM, Csernus VJ, Groot K, Kineman RD, Robinson WR, Schally AV (1999). Expression of growth hormone-releasing hormone (GHRH) messenger ribonucleic acid and the presence of biologically active GHRH in human breast, endometrial, and ovarian cancers. J Clin Endocrinol Metab. 84(2): 582-589. PubMed PMID: 10022420.
- 36 Kaulsay KK, Mertani HC, Törnell J, Morel G, Lee KO, Lobie PE (1999). Autocrine stimulation of human mammary carcinoma cell proliferation by human growth hormone. Exp Cell Res. 250(1): 35-50. PubMed PMID: 10388519.
- 37 Khandwala HM, McCutcheon IE, Flyvbjerg A, Friend KE (2000). The Effects of Insulin-like Growth Factors on Tumorigenesis and Neoplastic Growth. Endocrine Rev 21(3): 215–244. doi:10.1210/edrv.21.3.0399.
- 38 Kleinberg DL (1997). Early mammary development: growth hormone and IGF-1. J Mammary Gland Biol Neoplasia. **2**(1): 49-57. Review. PubMed PMID: 10887519.
- 39 Kozak RW, Haskell JF, Greenstein LA, Rechler MM, Waldmann TA, Nissley SP (1987). Type I and II Insulin-like Growth Factor Receptors on Human Phytohemagglutinin-Activated T Lymphocytes. Cellular Immunol 109 (2): 318–331. http://www.ncbi.nlm.nih. gov/pubmed/2959373.
- 40 Kusano K, Tsutsumi Y, Dean J, Gavin M, Ma H, Silver M et al. (2007). Long-term stable expression of human growth hormone by rAAV promotes myocardial protection post-myocardial infarction. J Mol Cell Cardiol. 42(2): 390-399. Epub 2006 Dec 15. PubMed PMID: 17174322.
- 41 Laban C, Bustin SA, Jenkins PJ (2003). The GH-IGF-I axis and breast cancer. Trends Endocrinol Metab. **14**(1): 28-34. Review. PubMed PMID: 12475609.

- 42 Lawler J, Detmar M (2004). Tumor progression: the effects of thrombospondin-1 and -2.Int J Biochem Cell Biol. **36**(6): 1038-1045. Review. PubMed PMID: 15094119.
- 43 Le Roith D, Bondy C, Yakar S, Liu JL, Butler A (2001). The somatomedin hypothesis: 2001. Endocr Rev. 22(1): 53-74. Review. PubMed PMID: 11159816.
- 44 Lesniak MA, Hedo JA, Grunberger G, Marcus-Samuels B, Roth J, Gorden P (1987). Receptors for insulin and growth hormone on lymphoid cells. Methods Enzymol. **150**: 701-723. PubMed PMID: 2828829.
- 45 Lincoln DT, Sinowatz F, Temmim-Baker L, Baker HI, Kölle S, and Waters MJ (1998). Growth Hormone Receptor Expression in the Nucleus and Cytoplasm of Normal and Neoplastic Cells. Histochem Cell Biol. **109**(2): 141–59. http://www.ncbi.nlm.nih.gov/pubmed/9504775.
- 46 Lowe WL Jr, Roberts CT Jr, Lasky SR, LeRoith D (1987). Differential expression of alternative 5' untranslated regions in mRNAs encoding rat insulin-like growth factor I. Proc Natl Acad Sci U S A. 84(24): 8946-50. PubMed PMID: 3480521; PubMed Central PMCID: PMC299668.
- 47 Mertani HC, Garcia-Caballero T, Lambert A, Gérard F, Palayer C, Boutin JM et al. (1998). Cellular expression of growth hormone and prolactin receptors in human breast disorders. Int J Cancer. **79**(2): 202-211. PubMed PMID: 9583737.
- 48 Minuto F, Del Monte P, Barreca A, Fortini P, Cariola G, Catrambone G et al. (1986). Evidence for an increased somatomedin-C/insulin-like growth factor I content in primary human lung tumors. Cancer Res. **46**(2): 985-988. PubMed PMID: 3940658.
- 49 Mukhina S, Mertani HC, Guo K, Lee KO, Gluckman PD, Lobie PE (2004). Phenotypic conversion of human mammary carcinoma cells by autocrine human growth hormone. Proc Natl Acad Sci U S A. 101(42): 15166-15171. Epub 2004 Sep 7. PubMed PMID: 15353581; PubMed Central PMCID: PMC524067.
- 50 Murphy LJ, Bell GI, Friesen HG (1987). Growth Hormone Stimulates Sequential Induction of c-Myc and Insulin-like Growth Factor I Expression in Vivo. Endocrinol **120** (5):1806–1812. doi:10.1210/endo-120-5-1806.
- 51 Neglia JP, Friedman DL, Yasui Y, Mertens AC, Hammond S, Stovall M et al.(2001). Second malignant neoplasms in five-year survivors of childhood cancer: childhood cancer survivor study. J Natl Cancer Inst. **93**(8): 618-29. PubMed PMID: 11309438.
- 52 Norsa A, Martino V (2006). Somatostatin, retinoids, melatonin, vitamin D, bromocriptine, and cyclophosphamide in advanced non-small-cell lung cancer patients with low performance status. Cancer Biother Radiopharm. 21(1): 68-73. PubMed PMID: 16480333.
- 53 Perry JK, Mohankumar KM, Emerald BS, Mertani HC, Lobie PE (2008). The contribution of growth hormone to mammary neoplasia. J Mammary Gland Biol Neoplasia. 13(1): 131-145. doi: 10.1007/s10911-008-9070-z. Epub 2008 Feb 7. Review. PubMed PMID: 18253708; PubMed Central PMCID: PMC2665193.
- 54 Perry JK, Emerald BS, Mertani HC, Lobie PE (2006). The Oncogenic Potential of Growth Hormone. Growth Hormone & IGF Research **16** (5–6): 277–289. doi:10.1016/j.ghir.2006.09.006.
- 55 Pines A, Rozen P, Ron E, Gilat T (1985). Gastrointestinal Tumors in Acromegalic Patients. The American Journal of Gastroenterology **80**(4): 266–269. http://www.ncbi.nlm.nih.gov/pubmed/3984995.
- 56 Rabinovitch A, Quigley C, Rechler MM (1983). Growth hormone stimulates islet B-cell replication in neonatal rat pancreatic monolayer cultures. Diabetes. 32(4): 307-312. PubMed PMID: 6339303.
- 57 Ratner RE, Hare JW (1983). Association of Acromegaly and Chondrosarcoma. Southern Med J. **76**(9): 1181–1182. http://www.ncbi.nlm.nih.gov/pubmed/6612402.
- 58 Ron E, Gridley G, Hrubec Z, Page W, Arora S, Fraumeni JF (1991). Acromegaly and Gastrointestinal Cancer. Cancer. **68**(8): 1673–1677. http://www.ncbi.nlm.nih.gov/pubmed/1913507.
- 59 Rutter MM, Rose SR (2007). Long-Term Endocrine Sequelae of Childhood Cancer. Current Opinion in Pediatrics 19(4): 480–487. doi:10.1097/MOP.0b013e3282058b56.
- 60 Schally AV, Varga JL (2006). Antagonists of growth hormonereleasing hormone in oncology. Comb Chem High Throughput Screen. 9(3): 163-170. Review. PubMed PMID: 16533148.

- 61 Sinowatz F, Breipohl W, Waters MI, Lincoln D, Lobie PE, Amselgruber W (1991). Growth Hormone Receptor Expression in the Dunning R 3327 Prostatic Carcinoma of the Rat. Prostate. **19**(4): 273–278. http://www.ncbi.nlm.nih.gov/pubmed/1754517.
- 62 Sklar CA, Mertens AC, Mitby P, Occhiogrosso G, Qin J, Heller G, Yasui Y, Robison LL (2002). Risk of disease recurrence and second neoplasms in survivors of childhood cancer treated with growth hormone: a report from the Childhood Cancer Survivor Study. J Clin Endocrinol Metab. 87(7): 3136-3141. PubMed PMID: 12107213.
- 63 Sommers CL, Byers SW, Thompson EW, Torri JA, Gelmann EP (1994). Differentiation state and invasiveness of human breast cancer cell lines. Breast Cancer Res Treat. 31(2-3): 325-335. PubMed PMID: 7881109. http://www.ncbi.nlm.nih.gov/pubmed/7881109.
- 64 Stewart SA, Weinberg RA (2006). Telomeres: cancer to human aging. Annu Rev Cell Dev Biol. **22**: 531-557. Review. PubMed PMID: 16824017.
- 65 Swerdlow AJ, Higgins CD, Adlard P, Preece MA (2002). Risk of Cancer in Patients Treated with Human Pituitary Growth Hormone in the UK, 1959-85: A Cohort Study. Lancet. (London, England) **360**(9329): 273–277. http://www.ncbi.nlm.nih.gov/pubmed/12147369.
- 66 Thiery JP (2002). Epithelial-Mesenchymal Transitions in Tumour Progression. Nature Reviews. Cancer 2(6): 442–454. doi:10.1038/ nrr822
- 67 Todisco M, Casaccia P, Rossi N (2001). Cyclophosphamide plus somatostatin, bromocriptin, retinoids, melatonin and ACTH in the treatment of low-grade non-Hodgkin's lymphomas at advanced stage: results of a phase II trial. Cancer Biother Radiopharm. **16**(2):171-177. PubMed PMID: 11385964.
- 68 Todisco M (2009). Chronic lymphocytic leukemia: long-lasting remission with combination of cyclophosphamide, somatostatin, bromocriptine, retinoids, melatonin, and ACTH. Cancer Biother Radiopharm. 24(3):353-355. doi: 10.1089/cbr.2008.0570. PubMed PMID: 19538058.
- 69 Vacas E, Muñoz-Moreno L, Valenzuela PL, Prieto JC, Schally AV, Carmena MJ et al. (2016). Growth hormone-releasing hormone induced transactivation of epidermal growth factor receptor in human triple-negative breast cancer cells. Peptides. 86:153-161. doi: 10.1016/j.peptides.2016.11.004. Epub 2016 Nov 2. PubMed PMID: 27816751.
- 70 Ward HC, Halliday D, Sim AJ (1987). Protein and Energy Metabolism with Biosynthetic Human Growth Hormone after Gastrointestinal Surgery. Annals Surgery 206(1): 56–61. http://www.ncbi.nlm.nih.gov/pubmed/3606231.
- 71 Wu X, Wan M, Li G, Xu Z, Chen C, Liu F et al. (2006). Growth hormone receptor overexpression predicts response of rectal cancers to pre-operative radiotherapy. Eur J Cancer. **42**(7): 888-894. Epub 2006 Mar 3. PubMed PMID: 16516462.
- 72 Wu ZS, Yang K, Wan Y, Qian PX, Perry JK, Chiesa J, et al. (2011). Tumor expression of human growth hormone and human prolactin predict a worse survival outcome in patients with mammary or endometrial carcinoma. J Clin Endocrinol Metab. 96(10): E1619-1629. doi: 10.1210/jc.2011-1245. Epub 2011 Aug 17. PubMed PMID: 21849525.
- 73 Yakar S, Leroith D, Brodt P (2005). The role of the growth hormone/insulin-like growth factor axis in tumor growth and progression: Lessons from animal models. Cytokine Growth Factor Rev. **16**(4-5): 407-420. Review. PubMed PMID: 15886048.
- 74 Yee D, Paik S, Lebovic GS, Marcus RR, Favoni RE, Cullen KJ et al. (1989). Analysis of insulin-like growth factor I gene expression in malignancy: evidence for a paracrine role in human breast cancer. Mol Endocrinol. 3(3): 509-517. PubMed PMID: 2747657.
- 75 Zhu T, Goh EL, Graichen R, Ling L, Lobie PE (2001). Signal transduction via the growth hormone receptor. Cell Signal. 13(9): 599-616. Review. PubMed PMID: 11495718.
- 76 Zhu Z, Mukhina S, Zhu T, Mertani HC, Lee KO, Lobie PE (2005). p44/42 MAP kinase-dependent regulation of catalase by autocrine human growth hormone protects human mammary carcinoma cells from oxidative stress-induced apoptosis. Oncogene. 24(23): 3774-3785. PubMed PMID: 15782123.
- 77 Ziel FH, Peters AL (1988). Acromegaly and gastrointestinal adenocarcinomas. Ann Intern Med. Sep 15;109(6):514-5. PubMed PMID: 3415113.